

Library Preparation

Illumina methods for library preparation include capture-based target enrichment and amplicon generation (Figure 3). With target enrichment, specific regions of interest are captured by hybridization to biotinylated probes, then isolated by magnetic pull-down. This highly multiplexed approach enables a wide range of applications for the discovery, validation, or screening of somatic variants. The TruSight RNA Pan-Cancer panel uses this method for library preparation.

Amplicon generation involves 2 submethods depending on the product chosen (Figure 3). TruSeq Custom Amplicon Low Input and TruSight Myeloid employ a hybridization-extension-ligation approach, creating a single strand template from a double-stranded DNA population that is later amplified via PCR. TruSight Tumor 15 utilizes a multiplexed PCR approach, amplifying the predefined targeted regions from genomic DNA. Sequencing of resulting amplicons from either method is useful for the discovery of rare somatic mutations in complex samples such as heterogenous tumors mixed with germline DNA.¹⁹

The TruSeq Custom Amplicon Low Input Dual Strand approach is an additional feature useful for somatic tumor profiling. This approach creates a mirror amplicon to the original amplicon generated during library preparation. This virtually reduces all idiosyncratic and systemic “noise” resulting from FFPE deamination, storage oxidation, and any other artifacts that arise in the handling, preparation, and resequencing process. The resulting DNA interrogation provides an accurate picture in profiling analysis.

Custom Panel Design

The TruSeq Custom Amplicon Low Input assay empowers researchers to create a custom panel targeting genes and regions of interest using Illumina DesignStudio™ software, a free, easy-to-use, online tool that provides optimized coverage (Figure 4). DesignStudio produces TruSeq Custom Amplicon Low Input probes with an average of > 94% *in silico* coverage across all gene sets. Illumina Concierge services offer support for optimization of probe design, functional evaluation and optimization of custom panels, and increasing target coverage.[†]

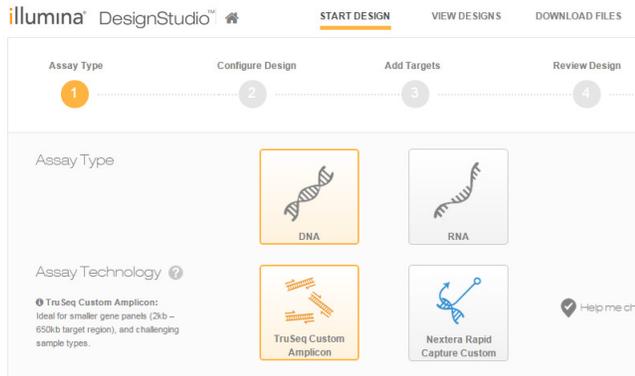


Figure 4: Custom Probe Design—Researchers can use DesignStudio to visualize targeted genomic regions and attempted amplicons to assess design coverage and score.

† For more information on Illumina Concierge service, contact an Illumina representative.

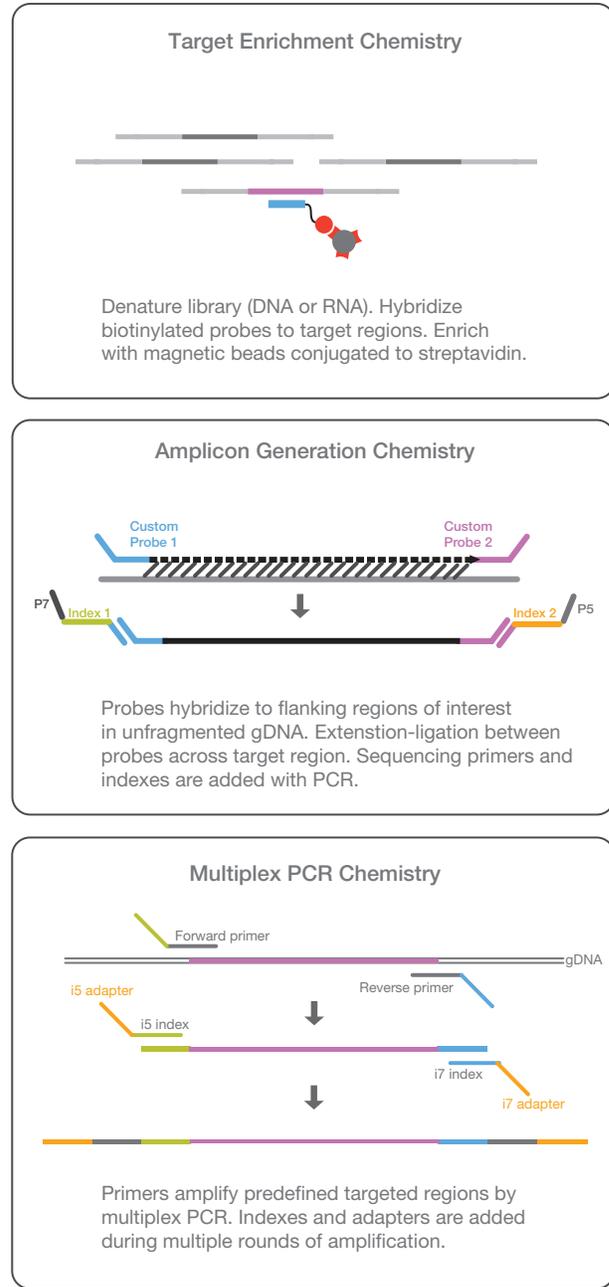


Figure 3: Library Preparation Methods—Illumina methods for sequencing library preparation include targeted enrichment, amplicon generation, and multiplex PCR.

Sequencing on the MiniSeq System

Whether using target enrichment or amplicon generation methods for library preparation, after sample libraries are prepared they can be easily sequenced on the MiniSeq System (Figure 5). It integrates clonal amplification and sequencing into a fully automated process on a single instrument. This eliminates the need to purchase and operate expensive, specialized equipment.

The MiniSeq System features load-and-go operation and an intuitive user interface that provides simple, step-by-step guidance through each stage of the sequencing run. It takes less than 5 minutes to load and set up a MiniSeq System. Sequencing runs can be completed in < 24 hours. MiniSeq reagent kits are available in Mid-Output and High-Output formats, allowing optimization of study designs based on read-length, sample number, and output requirements.



Figure 5: MiniSeq System—The MiniSeq System harnesses the latest advances in SBS chemistry and an easy, integrated workflow.

Simplified Data Analysis and Bioinformatics

Data analysis with the MiniSeq System requires no informatics expertise or command-line experience. It features Local Run Manager software, an onboard system for creating a run, monitoring status, and

automated sequencing data analysis post-run. Local Run Manager features a modular design that allows users to install and update individual analysis modules as needed, which generate simple reports for various sequencing applications.

In addition, sequencing data generated with the MiniSeq System can be instantly transferred, stored, and analyzed in the BaseSpace Computing Environment (Cloud-based or Onsite). BaseSpace Applications (Apps) provide expert-preferred data analysis tools in an intuitive, click-and-go user interface designed for informatics novices (Figure 6). These Apps support a range of common sequencing data analysis needs such as alignment, variant calling, and more. The BaseSpace ecosystem provides one of the largest collections of commercial and open-source analysis tools currently available. VariantStudio enables rapid filtering, identification, and annotation of disease-associated variants in flexible, structured reports (Figure 7).

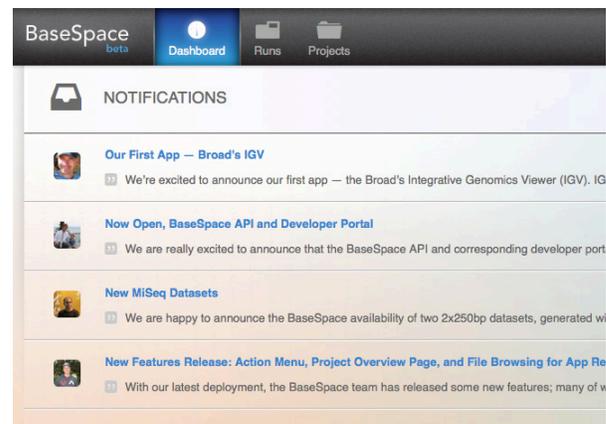


Figure 6: BaseSpace Dashboard—The BaseSpace Environment features an intuitive, click-and-go user interface to empower any researcher to perform their own informatics.

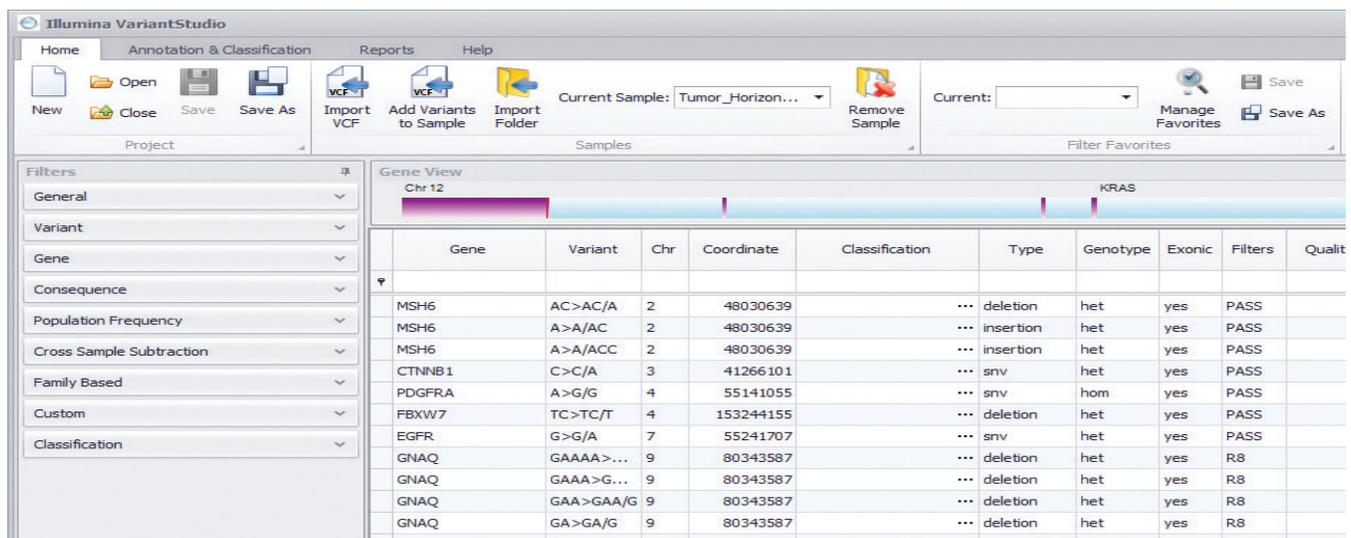


Figure 7: VariantStudio—VariantStudio software features an intuitive user interface that enables easy data analysis and exploration, without requiring informatics expertise. It aggregates information from a broad range of sources into a single database for comprehensive annotation of genomic data. Flexible report generation summarizes and annotates results.

Demonstrated Workflow – TruSight Tumor 15

Library Preparation

The TruSight Tumor 15 library preparation method enables multiplex PCR, which produces higher coverage uniformity and reduces the presence of primer dimers and FFPE-induced artifacts. This results in high accuracy and sensitivity for somatic variant analysis.²⁰

The TruSight Tumor 15 Protocol Guide is an easy-to-follow protocol for preparing DNA sequencing libraries, including DNA extraction, quantification, and in-process qualification steps. It leads users through each step of library preparation, listing necessary reagents and indicating safe stopping points.

Sample libraries were prepared from 20 ng of total input DNA following this protocol. Sequencing data sets were generated from 3 DNA controls of known variant compositions and 10 FFPE-extracted DNA samples from lung, colon, melanoma, and breast tumors previously characterized using the MiSeq® System.

Sequencing on the MiniSeq System

TruSight Tumor 15 sample library pools consisted of 8 samples per High-Output run (16 total; mix of DNA controls and FFPE-extracted tumor samples). Libraries were loaded onto the MiniSeq instrument along with the reagent cartridge and flow cell. Automated cluster generation and paired-end sequencing with a 300-cycle read was set up with Local Run Manager and carried out without any further user intervention, targeting 97% of bases at 500x coverage and taking 24 hours.

Data Analysis

Primary analysis (image analysis, base calling) was performed on the MiniSeq System. Additional analysis (demultiplexing, alignment, and variant calling) was performed with the TruSight Tumor 15 Local Run Manager Module (Figure 8) and VariantStudio.

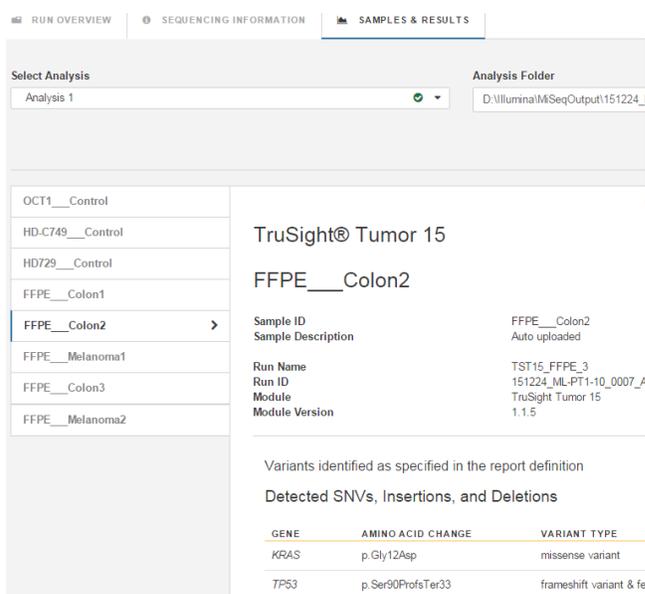


Figure 8: Local Run Manager—Local Run Manager software allows users to create a sequencing run, monitor status, and view results. Onboard data analysis is automatically performed upon run completion.

Results and Discussion

Running the TruSight Tumor 15 sequencing panel on the MiniSeq System achieves at least 95% of bases covered at ≥ 500x, which gives confidence in variant calling (Table 1). It enables detection of variants down to 1% (Table 2). TruSight Tumor 15 run on the MiniSeq System enables variant detection in many different sample types, including low quality FFPE samples (Table 3). Moreover, data generated on the MiniSeq system shows 100% concordance with previously characterized FFPE samples.

Table 1: TruSight Tumor 15 Coverage

Sample ID	Quality	% of Bases ≥ 500x	Amplicon Mean Coverage
FFPE_Colon1	Medium	99.7%	24,219x
FFPE_Colon2	Low	99.9%	20,763x
FFPE_Colon3	Low	99.2%	35,270x
FFPE_Colon4	High	100.0%	18,357x
FFPE_Colon5	High	100.0%	15,769x
FFPE_Melanoma1	Medium	99.7%	32,707x
FFPE_Melanoma2	Low	99.1%	41,640x
FFPE_Melanoma3	High	100.0%	17,285x
FFPE_Melanoma4	Low	95.7%	10,177x
FFPE_Breast1	High	99.1%	15,501x

